

US Veterinary Immune Reagent Network - Progress report for all species

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Abstract

The gene cloning phase of the project was completed during the early part of Year 3 (Spring 2008). In total, the Network cloned more than 150 complete expressed sequences of genes for cytokines, chemokines and cell surface markers of hematopoietic cells of cattle, horses, pigs, trout, catfish and chickens. The genes for chemokines and cytokines are being expressed at Kingfisher Biotech (KF) in yeast (Table 1) and in mammalian cells at Cornell for cell surface molecules as well as in the fish labs in bacteria or mammalian cells (Table 2). Many of these cytokines and chemokines have been expressed and assessment of bioactivity of chemokines and cytokines has been underway during Year 3 in the species labs for bovine, equine, chicken and swine (see Table 1). Fish cytokines are being produced starting in early 2009 (see schedule, Table 1). Cell surface molecule expression has continued since Year 1 and will be completed in Year 4. Generation of mAb to those cell surface molecules (and to the chemokines and cytokines selected) has made good progress during Year 3 and will continue through year 4 (Table 3). Additional information of the progress is summarized by species at www.vetimm.org on the various web pages and in attached PDFs on the web page as FASTA files, alignments of genes being expressed, commercially available reagents for the species, protocols for bioassays of cytokines and chemokines, primary references for the gene cloning and cytokine detection for each species.

Chemokines & Cytokines (Table 1). Following the completion of the gene cloning phase for the cytokines and chemokines, a large scale alignment and annotation of the expressed sequences was done to confirm the correctness of the sequences before expression at KF. These are shown on the website. Whole sequence included signal/leader sequence while those designated as KF are ready for expression in the yeast system in most cases without signal sequence or pro-peptides.

- ❖ All these genes are available for distribution to the scientific community upon request and we are investigating feasibility of deposition in gene banks.
- ❖ Virtually all sequences have been deposited in GenBank.
- ❖ Kingfisher (KF) biotech is in the process of expressing all these in *Pichia* and those expressed have been sent the various species coordinators for biotesting. This began in Feb 2008.
- ❖ The bioactive proteins are commercially available (see Kingfisher website www.KingfisherBiotech.com) and a limited amount directly distributed to scientists.
- ❖ A selected number will be used for producing mAbs by US-VIRN (see Table 2).

Cell Surface Molecules (Table 2) These genes were largely cloned by US-VIRN and are being transfected into CHO cells at Cornell by Bettina Wagner while the fish labs have produced their cell surface proteins in bacteria or mammalian cells; All will used for production of MAb.

- > Gene sequences have been/will be deposited into GenBank.
- > Most genes are available upon request.
- > All the proteins in this category have been, are in the process of or will be used for producing mAbs.
- > Transfected cell lines will also be available for testing other potential cross-reactive mAbs.

Monoclonal antibodies (Table 3) The cell surface molecules being expressed at Cornell in the Wagner Lab or in the Hansen or Mississippi fish labs will all be used for producing monoclonal antibodies (mAb) at Umass and Cornell or in the fish labs. In addition, a limited number of mAb will be made in this round against cytokines or chemokines that have shown bioactivity in the Species labs.

- ✓ These are being continually re-prioritized by the species coordinators in conjunction with the scientists in field but the current version is shown.
- ✓ The hybridoma cell lines that produce the mAbs will be deposited in cell banks for distribution upon request and distributed to a limited number of scientists directly by Network members; the mAb they produce will be available through commercial companies including Kingfisher.

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Table 2. CELL SURFACE MOLECULE PRODUCTION SCHEDULE

Expression in CHO cells: (-done; Dates - when protein will ready for mAb production) Those expressed bacteria are indicated as such	Molecule	Gene cloned*	Gene in expression vector	Stable transfectant	Protein purification	MAb in progress or made
Pig						
√ (recloning May '09 into different vector)	TCRa					
√ (recloning May '09 into different vector)	TCRβ					
	IL-4Rα					
	IL-13Rα1					
√ (peptide)	CD45					
Horse						
	CD40					
	CD23					
	CD25					
	CD28					
	FcRαR1					
July '09	IgD					
June '09	TCRa					
June '09	TCRβ					
June '09	TCRγ					
Bovine						
Feb '09	TCRδ					
March '09	IL-23R					
March '09	IL-10β					
March '09	CCR7					
Chicken						
June '09	CD80					
June '09	CD83					
June '09	CD86					
March '09	IL-2Rα					
May '09	CXCR4					
Catfish						
	TCRa					
	TCRβ					
	TCRγ					
	TCRδ					
√ (recloning July '09 into different vector)	TCRβ					
June '09	TCRδ					
√ in bacteria	IgD					
√ in bacteria	IgLα					
√ in bacteria	IgLδ					
Express in bacteria	β2-micro					
√ in bacteria	CD4L1					
	CD8 to be cloned					
Trout						
August '09	TCRa					
July '09	TCRβ					
June '09	TCRγ					
√ @ USGS	Blimp					
√ in bacteria	CD3g/d					
√ in bacteria	CD4					
√ in bacteria	CD4-REL					
√ in bacteria	CD8					
Express in bacteria	CD28					X
Express in bacteria	CD79A					
@ USGS	CD79B					
Express in bacteria	CD83					
Express in bacteria	CTLA4					
Express in bacteria	IgD					
Express in bacteria	IgT					
Express in bacteria	MHC IA (UBA)					X
@ USGS	MHC IIB					X
√ in bacteria	p56LCK					
√ in bacteria	Pax5					

All in the Table 2 will be used to produce mAbs as scheduled below in the central labs at Umass and Cornell or directly in Species labs if not indicated on this schedule

Those indicated by 'Yes' in Table 1 are slated for mAb production in this round as scheduled in table below;others will be targeted if re-funded

Table 1. CYTOKINE & CHEMOKINE PRODUCTION SCHEDULE

CATFISH	US-VIRN clone's GenBank #	Transformed in expression vector	Expressed in yeast (expected date)	Purified on FPLC (expected date)	Bioactivity affirmed	Will monoclonal antibodies be produced?
IFN γ	DQ124250	1/09	4/09			Yes
Type II IFN	AY847295	1/09	4/09			

CATTLE	US-VIRN clone's GenBank #	Transformed in expression vector	Expressed in yeast (expected date)	Purified on FPLC (expected date)	Bioactivity affirmed	Will monoclonal antibodies be produced?
CCL2	EU276069	√	√			
CCL5	EU276060	√	2/09			
CCL11	EU744565	√	√			
CXCL9	EU276061	√	12/08			
CXCL10	EU276062	√	2/09			
CXCL11	EU276063	√	√	√	YES	
IFN γ	EU276066	√	√	√	YES	
IFN α	EU276064	√	√	√	YES	yes
IFN β	EU276065	√	√	√		
IL-1 β	EU276067	√	√	√		
IL-2	EU276068	√	12/08			
IL-4	EU276069	√	√	√	YES	
IL-5	EU276070	√	3/09			
IL-6	EU276071	√	√			yes
IL-7	EU276072	√	3/09			
IL-8	EU276073	√	√ (additional 1/09)			
IL-10	EU276074	√	√	√		
IL-12p35	EU276075	√	2/09			
IL-12p40	EU276076	√	2/09			
IL-13	EU276077	√	√	√	YES	yes
IL-15	EU982380	√	√	√		
IL-17	EU982381	√	√	√		yes
IL-18	EU276078	√	√ (additional 1/09)			
IL-23	EU616677	√	1/09			yes
TNF α	EU276079	√	√ (additional 1/09)			

HORSE	US-VIRN clone's GenBank #	Transformed in expression vector	Expressed in yeast (expected date)	Purified on FPLC (expected date)	Bioactivity affirmed	Will monoclonal antibodies be produced?
CCL2	EU438774	√	1/09			
CCL3	EU438775	√	3/09			
CCL5	EU744564	√	3/09			
CCL11	In progress		3/09			
CXCL9	EU438776	√	2/09			
CXCL10	EU438777	√	2/09			
GM-CSF	EU438778	√	√ (additional 1/09)			
IFN γ	U04050	√	√			
IFN α	EU682378	√	√			
IL-1 β	EU438767	√	1/09			
IL-2	EU438768	√	√	√	YES	Yes
IL-4	EU438769	√	√	√	YES	Yes
IL-5	U91947	√	3/09			
IL-6	EU438770	√	√			Yes
IL-10	EU438771	√	3/09			
IL-12 p35/p40			4/09			
IL-13	EF645663	√	4/09			
IL-15	EU682379	√	12/08			
IL-17	EU744563-no s.s.	√	1/09			
IL-18	EU438772	√	√			
IL-23	EU438773	√	12/08			
TGF β	X99438	√	4/09			
TNF α	EU438779	√	2/09			

SWINE	US-VIRN clone's GenBank #	Transformed in expression vector	Expressed in yeast (expected date)	Purified on FPLC (expected date)	Bioactivity affirmed	Will monoclonal antibodies be produced?
CCL2	EU682382	√	√			Yes
CCL3L1	EU364893	√	3/09			
CCL4	EU364894	√	1/09			
CCL5	EU44561	√	√			
CXCL9	EU36897	√	2/09			Yes
CXCL10	EU364896	√	12/08		YES	Yes
CXCL11	EU682377	√	√	√	YES	Yes
IL-7	EU364895	√	√			
IL-13	EU682383	√	√ (repeat 12/08)	√	YES	Yes
IL-15	NM 214390	√	12/08			
IFN α	EU364896	√	√			
IFN- β 1	EU744582	√	√ (additional 1/09)			Yes
TNF-g	EU682384	√	√			

TROUT	US-VIRN clone's GenBank #	Transformed in expression vector	Expressed in yeast (expected date)	Purified on FPLC (expected date)	Bioactivity affirmed	Will monoclonal antibodies be produced?
IFN1	AM489418	√	12/08			Yes
IFN2	AJ682754	√	12/08			Yes
IFN3	AM236738	√	12/08			Yes
IFN γ	AJ616215	√	12/08			Yes

Table 3. MONOCLONAL ANTIBODY PRODUCTION SCHEDULE

Year	MONTH	University of Massachusetts				Cornell
		Spot 1	Spot 2	Spot 3	Spot 4	Spot 5
07	Jan	(Eq IL-8 - failed)				
08	Jan	(Eq IL-8 - failed)				Eq CD25
08	May	(Eq IL-8 - failed)				Sw IL-4R α
08	Sept	Catfish TCR γ	Catfish TCR α	Bo TCR γ	Eq CD40	Sw TCR β
08	Oct	Ch IL-10	Ch IL-16	Sw CD45	(Ch IL-10-2 nd)	Eq CD23
08	Nov	(Sw CD45-2 nd)	Catfish CD4L1	Catfish IgL δ	Catfish IgL λ	Eq CD28
08	Dec	Bo IL-13	Bo IL-23	Sw CXCL10	Sw CXCL11	Eq IL-4
09	Jan	Tr IFN1	Bo IFN α	Sw IFN β	Sw IL-13	Sw CCL2
09	Feb	Bo TCR δ	Ch IL-4	Ch IL-17	Bo IL-17	Sw IL-13R α
09	Mar	Bo IL-10 β	Tr IFN2	Ch IL-2R α	Catfish β 2-micro	Eq Fc ϵ R1 α
09	April	Tr IFN-3	Ch IL-17D	Bo CCR7		Eq IL-2
09	May	Ch CXCR4	Tr IFN-4	Bo IL-6	Tr CD3	Eq IL-6
09	June	Tr TCR γ	Ch CD80	Ch CD83	Ch CD86	Sw TCR β
09	July	Tr TCR α	Catfish TCR δ	Catfish TCR β		Eq TCR α
09	Aug	Tr TCR β	Tr CD4REL	Bo TCR α **	Ch IL-18	Sw TCR α
09	Sept	Catfish Type II IFN γ	Tr CD28		Ch IL-15	Eq TCR γ
09	Oct	Catfish TCR β 2	BOVINE*	SWINE*		Eq TCR δ
09	Nov	Catfish IgD	Tr CTLA4	SWINE*	Tr CD83	Eq IgD
09	Dec	Catfish CD8	Bo IL-23R			